

Lipofuscein as marker or player in accelerated cerebrovascular ageing in epilepsy?

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Skill & materials: Two-photon laser scanning microscopy (TPLSM); Super-resolution microscopy (STED); Cell culture and differentiation; labeling; human epileptic brain tissue

Key words/sectors: 前沿技□ / Frontier Technologies; 生物技□ / Biotechnology; 基□ 研究 / Basic Research; 基础研究 / Basic Research; 人类健康与疾病的生物学基□ / Biological Foundations of Human Health and Diseases

Basic Hypothesis: Mitochondrial activity and LPF production in young epilepsy patients and healthy elderly are comparable.

Cells with mitochondrial dysfunction undergo increased oxidative stress, causing difficulties in degrading proteins and lipids, ultimately aggregating in autofluorescent lipofuscein (LPF) droplets. As mitochondrial functioning decreases during ageing, LPF is often seen as “ageing” product. In the last 5 years, 2 expertise groups, a clinical group on epilepsy surgery¹⁻⁴ and a research group on advanced optical microscopy⁵⁻⁷, joined forces. Using microscopy and cutting-edge image analysis, we found LPF in pial arteries of young epilepsy (± 20 years) patients, at levels that are normally seen in healthy elderly (>65 years)^{8,9}. We are currently undertaking studies into the role of both microvascular changes^{10,11} and mitochondrial mechanisms underlying increased LPF distribution in the development of epilepsy.

We are looking for a talented and dedicated PhD student to carry out, analyze, and interpret cell culture studies with cells from 1) healthy individuals, 2) patients suffering from non-treatable epilepsy, and 3) patients suffering from mitochondrial dysfunction syndromes (POLG1, ATP6, tRNA(Leu)). Fibroblasts, stem cells, smooth muscle cells, and pericytes will be analyzed before and after metabolic challenge (glucose/galactose replacement). Imaging of mitochondrial activity and network, sub-mitochondrial structure and DNA changes, and LPF production (autofluorescence) are essential. We will compare these aspects to their readouts in brain pial arteries and tissues from control and epileptic individuals. Tissues are obtained during the surgery these patients undergo to decrease symptoms.

The collaboration between University Hospital and Imaging laboratories offers unique possibilities for the candidate to work on the cutting edge of two disciplines, i.e., imaging and resective brain surgery.

1. R. Haeren, S. Hartmans, J. De Mey, G. Hoogland, J. Dings, O. Schijns, J. van Overbeeke, and **K. Rijkers**. Cerebral Artery Vasoconstriction is Endothelin-1 Dependent Requiring Neurogenic and Adrenergic Crosstalk. *Current Neurovascular Research* 2017, 14(4), 306-315 **IF=3.7**
2. R. Haeren, S. van de Ven, **M. van Zandvoort**, H. Vink, J. van Overbeeke, G. Hoogland, and K. Rijkers. *Current Neurovascular Research* 2016, 13(3), 249-260 **IF=3.7**
3. R. Haeren, **K. Rijkers**, G. Hoogland, O. Schijns, J. Dings, J. Staals, **M. Van Zandvoort**, J. Van Overbeeke, H. Vink. In vivo assessment of the human cerebral microcirculation and its glycocalyx: a technical report. *Journal of Neuroscience Methods* 2018 303:114-125 **IF=2.8**
4. R. Haeren, H. Vink, J. Staals, **M.A.M.J. van Zandvoort**, J. Dings, J.J. van Overbeeke, G. Hoogland, **K. Rijkers**, and O.E.M.G. Schijns. Protocol for intraoperative assessment of the human cerebrovascular glycocalyx. *BMJ Open* 2017, 7(1): e013954 **IF=2.5**
5. C. Kuppe, C. van Roeyen, K. Berger, N. Kabgani, M. Vogt, **M. van Zandvoort**, B. Smeets, J. Floege, H. Gröne, and M. Moeller. Investigations of glucocorticoid action in GN. *J Am Soc Neph* 2017, 28(5):1408-1420 **IF=9.3**
6. A. Gemmink, M. Bosma, H.J. Kuijpers, J. Hoeks, G. Schaart, **M. van Zandvoort**, P. Schrauwen, and M. Hesselink. Decoration of intramyocellular lipid droplets with PLIN5 modulates fasting-induced insulin resistance and lipotoxicity in humans. *Diabetologia* 2016, 59(5): 1040-1048 **IF=7.1**
7. A. Baleanu-Curaj, Z. Wu, T. Lammers, C. Weber, **M. van Zandvoort**, and F. Kiessling. Non-invasive molecular ultrasound monitoring of vessel healing after intravascular surgical procedures in a preclinical setup. *Arterioscler Thromb Vasc Biol* 2015, 35(6): 1366-1373 **IF=6.6**
8. K. Hakvoort, L. Otto, R. Haeren, **K. Rijkers**, G. Hoogland, O. Schijns, J. Dings, J. Van Overbeeke, **M. van Zandvoort**. Shedding light on human cerebral Lipofuscein: an explorative study on identification and quantification. *Journal of Comparative Neurology* 2021, 529(3): 605-615 **IF=3.2**

9. K. Hakvoort, L. Otto, R. Haeren, K. Rijkers, G. Hoogland, O. Schijns, J. Dings, J. Van Overbeeke, **M. van Zandvoort**. Lipofuscin in cerebral arteries and tissue of epilepsy patients. *Journal of Comparative Neurology* 2020, doi 10.1002/cne.24971 **IF=3.4**.
10. Luqman Khan, Rick van Lanen, Govert Hoogland, Olaf Schijns, Kim Rijkers, **Dimitrios Kapsokalyvas**, Marc van Zandvoort, and Roel Haeren. Two-Photon Imaging to Unravel the Pathomechanisms Associated with Epileptic Seizures: A Review. *Appl. Sci.* **2021**, 11, 2404 **IF=2.7**
11. R.H.G.J. van Lanen, S. Melchers, G. Hoogland, I.E.M.G. Schijns, H. Vink, **M.A.M.J. van Zandvoort**, R.H.L. Haeren, **K. Rijkers**. Microvascular changes associated with Temporal Lobe Epilepsy: a narrative review. *J. of Cerebral flow and metabolism* 20210 **IF=6.2**